Gut microbiota affects brain development and behavior

Running title: Gut microbiota and brain

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Key message
• The gut microbiota can alter a host’s brain development and behavior.
• Gut bacteria communicate with the brain via the microbiota-gut-brain axis.
• Fecal microbial transplantation is a promising treatment strategy for autism spectrum disorder.
Abstract

The gut covers a large surface area of the body and faces various external factors. The brain works in concert with commensal microbes in the gut to efficiently process the enormous amount of chemical signals that enter the gut every day. This review discusses: 1) evidence that gut bacteria can alter brain development and behavior, 2) mechanisms by which gut bacteria communicate with the brain, 3) preclinical and clinical studies demonstrating the impact of gut microbiota on autism spectrum disorder, and 4) variables worth consideration by future research on gut bacteria.

Key words: Gut-brain axis, Microbiota, Autism, Development, Brain
**Introduction**

Except for the brain, which is equipped with a solid barrier, microbes cover all surfaces of the human body. Approximately 95% of human microbes live within the gastrointestinal tract. This is unsurprising as the surface area of the intestinal lumen corresponds to the size of two tennis courts (400 m²).\(^1\) According to recently revised estimates, there are nearly 38 trillion bacterial cells in the human body, mainly within the colon.\(^2\) As they have lived in the human body for millions of years, there is reasonable doubt that they inevitably evolved with humans. Considerable research has revealed the impact of gut bacteria on their hosts. Gut bacteria detect various compounds that enter the body and notify the host in several ways. Furthermore, their presence affects the development of various organs, metabolic processes, and the immune system.

Meaningful studies recently examined the microbiota-gut-brain axis hypothesis to explain the effect of the gut microbiota on the brain. This review will focus on: 1) interesting evidence that gut bacteria can alter host development and behavior even in the brain, a mighty fortress; 2) mechanisms by which gut bacteria communicate with the brain; 3) preclinical and clinical studies that demonstrate the impact of the gut microbiota on autism spectrum disorder (ASD); and 4) variables worth consideration in future research on gut bacteria.

**Evidence that the human brain is influenced by gut bacteria**

The blood-brain barrier (BBB) is impenetrable to all substances except for the limited nutrients the brain requires. Nevertheless, evidence suggests that the brain is influenced by the gut microbiota.

1. **Effect on neural development**
Germ-free (GF) mice exhibit increased adult hippocampal neurogenesis compared with control mice, an effect that occurs only in the dorsal hippocampus. The dorsal hippocampus plays a critical role in spatial learning and memory. There is also a crucial early life period in which microbiota colonization affects adult hippocampal neurogenesis.

Second, genes related to myelination and myelin plasticity are upregulated in the prefrontal cortex of GF mice. Re-colonization with conventional microbiota could reverse these changes in the myelin as well as activity-related gene expression. The prefrontal cortex is affected by neuropsychiatric disorders, such as attention deficit hyperactivity disorder, ASD, depression, and schizophrenia. Therefore, the link between the gut bacteria and these diseases is worth investigating.

Third, the expression of synaptic plasticity–related genes is altered in GF mice. Synaptophysin is a synaptic vesicle glycoprotein expressed by most neurons and neuroendocrine cells and an indirect marker of synaptic plasticity in the brain. The gut bacteria regulate synaptophysin expression and post-synaptic density-95, which is involved in excitatory synapse maturation.

Fourth, the gut microbiota is involved in the development of microglia, the primary immune cells in the brain. Microglia have many other functions in brain development including synaptic patterning, cell genesis, myelination, cell positioning, cell survival, axon dynamics, and cellular phagocytosis. Disturbances in the gut microbial community influence microglial development. GF or antibiotic-treated mice exhibit alterations in the microglial ratio and an immature phenotype. However, defective microglia are restored by replenishing the gut microbiota and short-chain fatty acids (SCFAs), bacterial fermentation products containing acetic propionic acid, and butyric acid. Microglial density in cortical specimens was normalized after addition of the SCFA mixture to the drinking water of GF mice for 4 weeks. Moreover, by analyzing transcriptional results in GF mouse microglia, Matcovitch-Natan et al.
identified the dysregulation of dozens of genes involved in microglial development, despite maturity.\(^7\)

Knowing that disturbances in the gut microbiota can lead to abnormal neurogenesis, it may be possible to manipulate the gut microbiota in brain diseases accompanied by abnormal myelin formation, synapses, or microglia.

### 2. Effect of gut microbiota on behavior or psychopathology

Gut microbial dysbiosis, which can occur at different stages of life, may contribute to the pathogenesis of various neuropsychiatric disorders and abnormal behaviors. The mechanisms by which gut bacteria communicate with the brain are as follows (Figure 1):

#### 1) Secretion of neurotransmitters and neuromodulators

Functioning neurotransmitters and neuromodulators can be isolated from gut bacteria (Table 1).

Monoamine neurotransmitters, such as dopamine, norepinephrine, and serotonin, can be derived from aromatic amino acids, such as phenylalanine, tyrosine, and tryptophan, by the action of aromatic amino acid decarboxylase within the gut bacteria. Gut bacterial species and enzymes involved in the metabolism of phenylalanine, tyrosine, and tryptophan were previously summarized by Liu et al.\(^25\) The altered expression of several neurotransmitters was observed in the central amygdala and dentate granule layer among the hippocampal subregions in GF mice.\(^26\) GF mice also exhibit anxiolytic behavior and increased motor activity, and turnover rates of noradrenaline, dopamine, and 5-hydroxytryptamine are significantly higher in the striatum.\(^5\) When GF mice are exposed to stress, anxious behaviors are more pronounced and the dopamine turnover rate in the upper brain involved in regulating stress and anxiety was
significantly altered. Some researchers have suggested that gut bacteria affect dopaminergic neurotransmission by modulating the mesocorticolimbic circuit.

Furthermore, the gut microbiota plays a critical role in central neurotrophin expression. Antibiotic-induced gut bacterial dysbiosis increases exploratory behavior and hippocampal brain-derived neurotrophic factor expression in mice. This was reversed by normalizing the gut microbiota.

Gut microbes affect the brain by directly secreting neurotransmitters and neuromodulators that act on the body or regulate their expression.

2) Proinflammatory cytokines

Alterations in cytokine levels that accompany microbial infections may affect the developing brain. Inflammatory cytokines can promote the conversion of progenitor cells into dopaminergic neurons and decrease dendritic development. Intrauterine exposure to specific gastrointestinal microbial pathogens can induce multiple psychopathologies, such as memory impairment or schizophrenia later in life. The impact of maternal infection on fetal neurodevelopment is expected to vary with gestational age. For example, a maternal infection in the first trimester of pregnancy increases the risk of schizophrenia in the offspring.

3) Enteric nervous system and vagus nerve

Although the vagus nerve can perform both efferent and afferent roles, approximately 80% of nerve fibers are sensory organs that are mainly responsible for transmitting information about the state of the body organs to the brain. Postprandial satiety and sedation are produced partially by the active vagal afferent nerves in response to food intake. Likewise, the gut microbiota can signal the enteric nervous system and send signals to the brain via the vagus nerve. Treatment
with *Lactobacillus rhamnosus* reduced stress-induced corticosterone levels and anxiety- and depression-related behaviors in rats.\(^{37}\) Notably, no neurochemical or behavioral effects were noted in mice after vagal nerve dissection, confirming the vagus nerve as the principal communication pathway between the gut bacteria and brain.

4) **Neuroactive metabolites**

The gut microbiota can modulate host behavior via their metabolites. The parietal cells of the colon produce most of the serotonin in the periphery (60% in rats, 90% in humans). Serotonin production and secretion are affected by microbial metabolites including indole, SCFAs, secondary bile acids, α-tocopherol, p-aminobenzoate, and tyramine.\(^{23}\) Furthermore, gut bacterial taxa and their metabolites differ between ASD and control mice.\(^{38}\) When ASD mice are fed specific amino acid metabolites produced by bacterial fermentation (taurine, 5-aminovaleric acid), behavioral abnormalities (repetitive behavior and impaired social communication) significantly improve.

Moreover, gut microbes ferment polysaccharides to produce SCFAs (usually sodium butyrate). Butyrate-producing bacterial taxa are less abundant in children with ASD than in typically developing children.\(^{39}\) Butyrate also strengthens the BBB by creating dense connections between neurons.\(^{40}\) We previously questioned the association between the gut microbiota and neuropsychiatric disorders accompanying BBB permeability.\(^{41}\) Evidence also suggests that gut bacterial metabolites play a role in hunger. Hunger can be modulated by glucagon-like peptide-1 secreted by colonic enteroendocrine L cells in response to the bacterial metabolite indole, which stimulates colonic vagal afferent activity in rats.\(^{42}\)

2. **Impact of gut microbiota on ASD: preclinical and clinical studies**

Many researchers have attempted to modify the gut microbiota in patients to treat various brain disorders. ASD is the most actively studied developmental disorder in this field. However, no
results have clearly indicated a specific bacterial strain responsible for ASD, as shown in the meta-analyses below. Xu et al. (2019) analyzed nine studies.\(^{43}\) They identified a lower abundance in the ASD groups in the *Akkermansia, Bacteroides, Bifidobacterium, Escherichia coli,* and *Enterococcus* genera and a greater abundance in the *Faecilobacterium, Ruminococcus,* and *Lactobacillus* genera. Iglesias-Vázquez et al.’s analysis of 18 studies assessing 493 children with ASD and 404 controls reported a lower abundance in children with ASD in the *Bifidobacterium* and *Coprococcus* genera and a greater abundance in the genera *Faecalibacterium, Bacteroides, Parabacteroides, Clostridium,* and *Phascolarctobacterium.*\(^{44}\) Andreo-Martínez et al. analyzed 18 studies that assessed 642 patients and 356 controls.\(^{45}\) The *Streptococcus* and *Bifidobacterium* genera were less abundant in children with ASD. The included studies used different assessment methods, which could have been confounding factors. A recently published Korean study also reported inconsistent results: lower *Bacteroides* levels and higher *Bifidobacterium* levels in ASD patients versus controls.\(^{46}\)

Nevertheless, several clinical trials have attempted to alter the gut microbiota to treat patients with ASD (Table 2). Of them, microbial transfer therapy for children with ASD showed promising results with a steady improvement in core autism symptoms.\(^{47}\) Moreover, the behavioral effects of the fecal microbiotal transplant persisted even at the 2-year follow-up.\(^{48}\) However, behavioral outcomes have been inconsistent among studies using probiotics or prebiotics.
Points to consider when designing a gut microbiota study

More clinical evidence is warranted to standardize treatments for manipulating the gut microbiota in the future. When planning a clinical study, factors that can influence the gut microbiota must be recognized.

Clinical research on the gut microbiota is especially difficult because of the many possible variables that can change the outcome. However, it is difficult to control them simultaneously. As we analyzed earlier, the inconsistent results of many clinical studies on gut microbiota may be due to poor control of these confounding factors. Many clinical studies have failed to incorporate these variables into their study design; moreover, whether they were unaware or deliberately ignored is unclear. Figure 2 shows several known factors that can alter the gut microbiota, including diet, medicine, age, delivery mode, stress, and host factors. For a successful clinical study, it is necessary to fully recognize and control these variables as much as possible.

Conclusions

The brain works in concert with commensal gut microbes to efficiently process the enormous amount of chemical signals that enter the gut daily. Elucidating the relationship between the gut microbiota and the brain has become essential to further our understanding of the brain’s development and behavior.

Footnotes

Conflicts of interest: No potential conflict of interest relevant to this article was reported.

Funding: This study received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

ORCID
References


Table 1. Neuroactive amines and amino acids released by the gut bacteria

<table>
<thead>
<tr>
<th>Neurochemicals</th>
<th>Genus</th>
<th>References</th>
</tr>
</thead>
</table>
| Glutamate      | *Corynebacterium glutamicum*, *Lactobacillus*  
                             *plantarum*, *Lactobacillus paracasei*, *Lactococcus lactis* | 8-10)      |
| GABA           | *Escherichia coli*, *Pseudomonas*, *Lactococcus*, *Lactobacillus*,  
                             *Bifidobacterium* | 11-15)     |
| Dopamine       | *Escherichia*, *Bacillus*, *Lactococcus*, *Lactobacillus*, *Streptococcus* | 16, 17)    |
| Norepinephrine | *Escherichia*, *Bacillus*                  | 17)        |
| Serotonin      | *Streptococcus*, *Escherichia*, *Enterococcus*, *Lactococcus*,  
                             *Lactobacillus*, *Corynebacterium* | 16-18)     |
| Histamine      | *Lactobacillus*, *Lactococcus*, *Streptococcus*, *Enterococcus* | 19-21)     |
| Acetylcholine  | *Lactobacillus*, *Bacillus*                | 22-24)     |
Table 2. Behavioral outcomes of clinical trials that engineered the gut microbiota in patients with autism spectrum disorder

<table>
<thead>
<tr>
<th>Intervention group</th>
<th>Reference</th>
<th>Population (age, yrs)</th>
<th>Intervention</th>
<th>Study Design</th>
<th>Behavioral outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Probiotics alone or in combination</td>
<td>Sandler et al. (2000)⁴⁷</td>
<td>11 regressive-onset ASD with antecedent antimicrobial use (3.5–7), 62 ASD (4–16)</td>
<td>Vancomycin + (Lact acidophilus, Lact bulgaricus, Bifid bifidum), Lact plantarum</td>
<td>Open-label trial, RDBPC</td>
<td>Improvements in ASD severity (↓ CARS), short-term</td>
</tr>
<tr>
<td></td>
<td>Parracho et al. (2010)⁴⁸</td>
<td></td>
<td></td>
<td></td>
<td>Decreased disruptive, anti-social behavior, anxiety, and communication disturbances</td>
</tr>
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<td></td>
<td>Kaluzna-Czaplinska e Blaszczyk (2012)⁴⁹</td>
<td>22 ASD with GI symptoms (4–10)</td>
<td>Lact acidophilus</td>
<td>Open-label trial</td>
<td>Improvement in the ability to concentrate and carry out orders</td>
</tr>
<tr>
<td></td>
<td>Arnold et al. (2019)⁵⁰</td>
<td>13 ASD with GI symptoms and anxiety (2–11)</td>
<td>4 Lact strains + 3 Bifid strains + 1 Strept strain</td>
<td>RDBPC, crossover</td>
<td>No significant behavioral change (PRAS-ASD, ABC, SRS)</td>
</tr>
<tr>
<td></td>
<td>Liu et al. (2019)⁵¹</td>
<td>80 ASD (7–15)</td>
<td>Lact plantarum</td>
<td>RDBPC</td>
<td>Improvement in some autism symptoms, primarily those associated with disruptive and rule-breaking behaviors and hyperactivity/impulsivity (more prominent in younger children)</td>
</tr>
<tr>
<td></td>
<td>Niu et al. (2019)⁵²</td>
<td>114 ASD (ABA vs. ABA + probiotics), 40 HC (3–8)</td>
<td>3 Bifid strains + 3 Lact strains</td>
<td>Open-label, two-arm, randomized trial</td>
<td>Improvements in ASD severity (↓ ATEC)</td>
</tr>
<tr>
<td></td>
<td>Santocchi et al. (2020)⁵³</td>
<td>85 ASD (mean, 4.2)</td>
<td>4 Lact strains + 3 Bifid strains + 1 Strept strain</td>
<td>RDBPC</td>
<td>Improvements in ASD severity (↓ ADOS-CS) in the ASD without GI symptoms, although not significant in the ASD vs. the placebo</td>
</tr>
<tr>
<td>Study Reference</td>
<td>Participants</td>
<td>Intervention</td>
<td>Study Design</td>
<td>Outcomes</td>
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<tr>
<td>Mensi et al. (2021)</td>
<td>131 ASD (mean, 86.1 ± 41.1 mo)</td>
<td>Lact Plantarum (105 ASD), Others (26 ASD)</td>
<td>Open-label trial</td>
<td>Improvements in ASD severity (↓ CGI); Greater improvements in the Lact Plantarum group, no difference depending on the presence of GI symptom</td>
<td></td>
</tr>
<tr>
<td>Shaaban et al. (2018)</td>
<td>30 ASD (5-9) 30 HC children (5–9)</td>
<td>Lact acidophilus + Lact rhamnosus + Bifid longum and dried carrot</td>
<td>Open-label trial</td>
<td>Improvements in ASD severity (↓ ATEC)</td>
<td></td>
</tr>
<tr>
<td>Sanctuary et al. (2019)</td>
<td>8 ASD with GI symptoms (2–11)</td>
<td>Bifidobacterium infantis + bovine colostrum product</td>
<td>Randomized double-blind trial, crossover</td>
<td>Improved some atypical behaviors (irritability, stereotypies, hyperactivity, lethargy)</td>
<td></td>
</tr>
<tr>
<td>Wang et al. (2020)</td>
<td>26 ASD (3–9)</td>
<td>Bifid infantis and lactis, Lact rhamnosus and paracasei + fructooligosaccharide</td>
<td>RDBPC</td>
<td>Improvements in ASD severity (↓ ATEC)</td>
<td></td>
</tr>
<tr>
<td>Grimaldi et al. (2018)</td>
<td>41 ASD (4–11)</td>
<td>Bimuno galactooligosaccharides</td>
<td>RDBPC</td>
<td>Improved only in anti-social behavior</td>
<td></td>
</tr>
<tr>
<td>Inoue et al. (2019)</td>
<td>13 ASD (4–9)</td>
<td>Partially hydrolyzed guar gum with β-endogalactomannase produced by a strain of Aspergillus niger</td>
<td>Open-label trial</td>
<td>Decreased behavioral irritability</td>
<td></td>
</tr>
<tr>
<td>Kang et al. (2017)</td>
<td>18 ASD with GI symptoms (7–16)</td>
<td>Bowel prep. with vancomycin + SHGM orally or rectally</td>
<td>Open-label trial</td>
<td>Improvements in ASD severity (↓ CARS, PGI-III, ABC, SRS)</td>
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</tr>
<tr>
<td>Kang et al. (2019)</td>
<td>18 ASD with GI symptoms (7–17)</td>
<td>2-year follow-up after SHGM orally or rectally</td>
<td>Open-label trial</td>
<td>only 17% were rated as severe ASD, 39% were in the mild to moderate, and 44% were below the ASD diagnostic cut-off scores. (83% of participants were severe ASD at the beginning of the trial.)</td>
<td></td>
</tr>
<tr>
<td>Li et al. (2021)</td>
<td>40 ASD with GI symptoms (mean, 8.03 ± 3.73) 16 HC</td>
<td>Bowel prep. with polyethylene glycol + cFM orally or rectally</td>
<td>Open-label trial</td>
<td>Improved mood, behavior, emotion, language, and core ASD symptoms (↓ CARS, ABC, SRS) Parent’s decreased anxiety levels</td>
<td></td>
</tr>
</tbody>
</table>
ABA, applied behavioral analysis; ABC, Aberrant Behavior Checklist; ADOS-CS, Autism Diagnostic Observation Schedule - Calibrated Severity; ASD, autism spectrum disorder; ATEC, Autism Treatment Evaluation Checklist; Bifid, *Bifidobacterium*; CARS, Childhood Autism Rating Scale; cFM, fecal microbiota-filled capsules; CGI, Clinical Global Impression; GI, gastrointestinal; HC, healthy controls; Lact, *Lactobacillus*; mo, months; PGI-III, Parent Global Impressions-III; PRAS-ASD, Parent-Rated Anxiety Scale for ASD; Prep, preparation; RDBP, randomized double-blind placebo-controlled; SHGM, standardized human gut microbiota; SRS, Social Responsiveness Scale; Strept, *Streptococcus*; yrs, years
**Figure legends**

Fig. 1. Microbiota-gut-brain axis

The mechanisms by which gut bacteria communicate with the brain include the secretion of neurotransmitters, neuromodulators, and proinflammatory cytokines; engaging the enteric nervous system and vagus nerve; and producing neuroactive metabolites

Fig. 2. Factors affecting gut microbiota composition
Factors affecting gut microbiota

- Diet
- Medicine
- Age
- Stress
- Mode of delivery
- Host factors
  - mucus, AMPs, IgA, miRNAs